

Thermodynamic study of the binding of calcium and magnesium ions with myelin basic protein using the extended solvation theory

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The interaction of myelin basic protein (MBP) from the bovine central nervous system with Ca^{2+} and Mg^{2+} ions, named as M^{2+} , was studied by isothermal titration calorimetry at 27 °C in aqueous solution. The extended solvation model was used to reproduce the enthalpies of $\text{MBP}+\text{M}^{2+}$ interactions. The solvation parameters recovered from the extended solvation model were attributed to the structural change of MBP due to the metal ion interaction. It was found that there is a set of two identical and noninteracting binding sites for Ca^{2+} and Mg^{2+} ions.

Keywords myelin basic protein; isothermal titration calorimetry; binding parameter

The energy of biochemical reactions or molecular interactions at constant temperature is measured by isothermal titration calorimetry (ITC) [1,2]. ITC gives invaluable information about biomacromolecule–ligand interaction. During the last two years, we attempted to study the metal ion binding on different proteins [3–5]. Metal ions change the conformational stability and formation of aggregates. The importance of metal ions such as Ca^{2+} and Mg^{2+} in regulating protein stability and function has been widely reported [5–10].

We have previously developed a theory to account for the solvation of solutes in mixed solvent systems. Studies within our group are aimed at developing an understanding of how the binding proteins of the metal ions and other ligands affect the stability of the biomolecules. One of the

unique aspects of our approach is studying the stability of proteins by using the extended solvation model. Myelin basic protein (MBP) is one of the most abundant proteins of the myelin sheath of the central nervous system (CNS), and its primary role is generally considered to be maintenance of the stability of the sheath by holding together the apposing cytoplasmic leaflets of the oligodendrocyte membrane [11]. Previous investigations have revealed that the efficiency of MBP in preserving the compactness and the stability of the myelin membrane appeared to be enhanced in the presence of zinc (Zn) ions. There is also evidence that Zn stabilizes the *in vitro* self association of MBP dissolved in a phosphate buffer [12]. Since MBP addition to its associations with lipids, it has been shown to interact with calmodulin and cytoskeletal proteins such as actin and tubulin. Thus, functional roles for MBP in phosphoinositide-mediated signal transduction and other processes have been postulated [13]. The *MBP* gene encodes two families of paired formation of myelin sheets in primary cultures of cortical proteins: the classic MBPs and the Golli proteins, the function of which is less well understood. Previous work has suggested that Golli proteins may play a role in Ca homeostasis in oligodendrocytes and in T-cells [14].

Ca and Mg are of great physiologic importance by their intervention in many enzymatic systems and their function in neural excitability, muscle contraction blood coagulation, bone formation, hormone secretion, and cell adhesion [15]. The human body is equipped with an efficient negative feedback system that counteracts variations of Ca and Mg balance [16]. The maintenance of the Ca and Mg balance is controlled by the concerted action of intestinal absorption, renal excretion, and exchange with bone. After years of research, rapid progress has been made recently in identification and characteriza-

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tion of the Ca and Mg transport proteins that contribute to the delicate balance of divalent cations [15]. The interactions of MBP with Ca^{2+} -calmodulin have been previously investigated by fluorescence spectroscopy [17]. The apparent associated equilibrium binding constant for MBP interaction with Ca^{2+} -calmodulin was previously reported to be $2.1 \pm 0.1 \mu\text{M}^{-1}$ [18]. MBP is an “intrinsically unstructured” or “natively unfolded” protein, therefore its three-dimensional structure might only be determined in its interaction with another protein [19–22]. As a clear understanding of operational stability constitutes an important goal in protein technology, our efforts aimed at elucidation of the structure-stability using the extended solvation model. This model is able to correlate the solvation parameters to the effect of metals on the stability of a protein in a very simple way.

The present paper reports some interesting experimental data for the heats of interaction of Ca^{2+} and Mg^{2+} ions with MBP, and analyzes these using the extended solvation theory for the first time. With regard to the importance of the presence of balance between Ca and Mg values in humans, and the role of MBP in Ca homeostasis in oligodendrocytes and T-cells, it seems that the present study has significant applications in pharmacology, neuroscience and drug delivery.

Materials and Methods

MBP from bovine CNS was obtained from Sigma (St. Louis, USA). Protein concentrations were determined from absorbance measurements at 277 nm in 1 cm quartz cuvette. All other materials and reagents were of analytical grade, and solutions were made in 50 mM NaCl using double-distilled water.

The isothermal titration microcalorimetric experiments were performed with the four-channel commercial microcalorimetric system, Thermal Activity Monitor 2277 (Thermometric, Jarfalla, Sweden). The titration vessel was made from stainless steel. The metal nitrate solution (2 mM) was injected, by use of a Hamilton syringe, into the calorimetric titration vessel, which contained 1.8 ml MBP (60 μM). Thin (0.15 mm inner diameter) stainless steel hypodermic needles, permanently fixed to the syringe, reached directly into the calorimetric vessel. Injection of metal nitrate solutions into the perfusion vessel was repeated 30 times, with 20 μl per injection. The calorimetric signal was measured by a digital voltmeter that was part of a computerized recording system. The heat of each injection was calculated by the “Thermometric Digitam 3” software program. The heats of dilution

of the metal ion solutions were measured as described above, except that MBP was excluded. The microcalorimeter was frequently calibrated electrically during the course of the study.

Results

The enthalpies interaction of myelin basic protein (MBP) with Ca^{2+} and Mg^{2+} ions, were calculated in $\text{kJ}\cdot\text{mol}^{-1}$. The results of MBP+ Ca^{2+} and MBP+ Mg^{2+} were shown in **Table 1** and **2** respectively.

Table 1 The enthalpies of MBP+ Ca^{2+} interactions

[Ca] _T (μM)	[MBP] _T (μM)	ΔH ($\text{kJ}\cdot\text{mol}^{-1}$)	ΔH_{dilut} ($\text{kJ}\cdot\text{mol}^{-1}$)
8.1967213	13.2786890	3.0946502	4.0866667
16.1290320	13.0645160	5.7078189	3.7666667
23.8095240	12.8571430	7.9218107	3.4800000
31.2500000	12.6562500	9.7942387	3.2133333
38.4615380	12.4615380	11.3909470	2.9880000
45.4545450	12.2727270	12.7530860	2.7888889
52.2388060	12.0895520	13.9423870	2.6133333
58.8235290	11.9117650	14.9588480	2.4550000
65.2173910	11.7391300	15.8477370	2.3125926
71.4285710	11.5714290	16.6255140	2.1853333
77.4647890	11.4084510	17.3127570	2.0678788
83.3333330	11.2500000	17.9259260	1.9622222
89.0410960	11.0958900	18.4732510	1.8666667
94.5945950	10.9459460	18.9629630	1.7795238
100.0000000	10.8000000	19.4032920	1.6995556
105.2631600	10.6578950	19.8065840	1.6266667
110.3896100	10.5194810	20.1687240	1.5596078
115.3846200	10.3846150	20.5020580	1.4977778
120.2531600	10.2531650	20.8065840	1.4403509
125.0000000	10.1250000	21.0864200	1.3866667
129.6296300	10.0000000	21.3456790	1.3365079
134.1463400	9.8780488	21.5843620	1.2896970
138.5542200	9.7590361	21.8065840	1.2466667
142.8571400	9.6428571	22.0123460	1.2058333
147.0588200	9.5294118	22.2057610	1.1674667
151.1627900	9.4186047	22.3868310	1.1317949
155.1724100	9.3103448	22.5555560	1.0982716
159.0909100	9.2045455	22.7119340	1.0666667
162.9213500	9.1011236	22.8600820	1.0367816
166.6666700	9.0000000	23.0000000	1.0084444

ΔH , and the enthalpies of dilution of $\text{Ca}(\text{NO}_3)_2$ solution in water; ΔH_{dilut} , against Ca^{2+} and MBP concentrations.

Table 2 The enthalpies of MBP+Mg²⁺ interactions

[Mg] _T (μM)	[MBP] _T (μM)	Δ <i>H</i> (kJ·mol ⁻¹)	Δ <i>H</i> _{dilut} (kJ·mol ⁻¹)
8.1967213	13.2786890	3.4197531	3.9400000
16.1290320	13.0645160	6.2962963	3.9400000
23.8095240	12.8571430	8.7037037	3.5511111
31.2500000	12.6562500	10.7242800	3.2516667
38.4615380	12.4615380	12.4279840	3.0053333
45.4545450	12.2727270	13.8683130	2.7788889
52.2388060	12.0895520	15.0987650	2.5866667
58.8235290	11.9117650	16.1563790	2.4175000
65.2173910	11.7391300	17.0699590	2.2711111
71.4285710	11.5714290	17.8641980	2.1333333
77.4647890	11.4084510	18.5637860	2.0139394
83.3333330	11.2500000	19.1810700	1.9061111
89.0410960	11.0958900	19.7283950	1.8097436
94.5945950	10.9459460	20.2181070	1.7200000
100.0000000	10.8000000	20.6543210	1.6395556
105.2631600	10.6578950	21.0493830	1.5654167
110.3896100	10.5194810	21.4115230	1.4980392
115.3846200	10.3846150	21.7366260	1.4359259
120.2531600	10.2531650	22.0329220	1.3785965
125.0000000	10.1250000	22.3086420	1.3256667
129.6296300	10.0000000	22.5596710	1.2765079
134.1463400	9.8780488	22.7901230	1.2309091
138.5542200	9.7590361	23.0041150	1.1889855
142.8571400	9.6428571	23.2057610	1.1491667
147.0588200	9.5294118	23.3909470	1.1120000
151.1627900	9.4186047	23.5637860	1.0771795
155.1724100	9.3103448	23.7242800	1.0444444
159.0909100	9.2045455	23.8765430	1.0130952
162.9213500	9.1011236	24.0164610	0.9836782
166.6666700	9.0000000	24.1522630	0.9555556

Δ*H*_{dilut} and the enthalpies of dilution of Ca(NO₃)₂ solution in water; Δ*H*_{dilut} against Ca²⁺ and MBP concentrations.

Discussion

We have shown previously that the enthalpies of the solute-solvent (M²⁺+MBP+water in this case) interactions in the aqueous solvent (M²⁺+water in the present case) system, can be accounted for quantitatively in terms of three factors: preferential solvation by the components of a mixed solvent, weakening or strengthening of solvent-solvent bonds by the solute and the change in the enthalpy of the solute-solvent interactions [23–31]. This treatment leads to:

$$\Delta H = \Delta H_{\max} x'_B - \delta_A^\theta (x'_A L_A + x'_B L_B) - (\delta_B^\theta - \delta_A^\theta) (x'_A L_A + x'_B L_B) x'_B \quad 1$$

The parameters δ_A^θ and δ_B^θ are the indexes of MBP stability in the low M²⁺ ion concentrations and in the maximum concentration of the M²⁺ upon saturation of all MBP respectively. Cooperative binding requires that the macromolecules have more than one binding site, since cooperativity results from the interactions between binding sites with ligands. If the binding of a ligand at one site increases the affinity for the ligand at another site, the macromolecule exhibits positive cooperativity. Conversely, if the binding of a ligand at one site lowers the affinity for a ligand at another site, the protein exhibits negative cooperativity. If the ligand binds at each site independently, the binding is non-cooperative: $p < 1$ or $p > 1$ indicate positive or negative cooperativity of the macromolecule for binding with the ligand, respectively; $p = 1$ indicates that the binding is non-cooperative. x'_B can be expressed as follow:

$$x'_B = \frac{px_B}{x_A + px_B} = \frac{v}{g} \quad 2$$

x_B is the fraction of the metal ion needed for saturation of the binding sites, and $x_A = 1 - x_B$ is the fraction of unbounded M²⁺ ions. Now the model is a simple mass action treatment, with metal ions replacing water molecules, at the binding sites in the present case. We can express x_B fractions, as the total M²⁺ concentrations divided by the maximum concentration of the M²⁺ upon saturation of all MBP as follows:

$$x_B = \frac{[M^{2+}]_T}{[M^{2+}]_{\max}}; \quad x_A = 1 - x_B \quad 3$$

$[M^{2+}]_T$ is the total concentration of metal ions and $[M^{2+}]_{\max}$ is the maximum concentration of the M²⁺ upon saturation of all MBP. In general, there will be “ g ” sites for binding of M²⁺ per MBP molecule and v is defined as the average moles of bound M²⁺ per mole of total MBP. L_A and L_B are the relative contributions of unbounded and bounded metal ions to the enthalpies of dilution in the absence of MBP and can be calculated from the enthalpies of dilution of M²⁺ in buffer, Δ*H*_{dilut}, as follow:

$$L_A = \Delta H_{\text{dilut}} + x_A \left(\frac{\partial \Delta H_{\text{dilut}}}{\partial x_B} \right); \quad L_B = \Delta H_{\text{dilut}} - x_A \left(\frac{\partial \Delta H_{\text{dilut}}}{\partial x_B} \right) \quad 4$$

The enthalpies of M²⁺+MBP interactions, Δ*H*, were fitted to **Equation 1** over the whole M²⁺ compositions. In the procedure the only adjustable parameter (p) was changed until the best agreement between the experimen-

tal and calculated data was approached (Figs. 1–3). δ_A^θ and δ_B^θ parameters have been also optimized to fit the data. The optimized δ_A^θ and δ_B^θ values are recovered from the coefficients of the second and third terms of **Equation 1**. The small relative standard coefficient errors and the high r^2 values (0.99999) support the method. The agreement between the calculated and experimental results (Fig. 1) is striking, and gives considerable support to the use of **Equation 1**.

Φ is the fraction of the MBP molecule undergoing com-

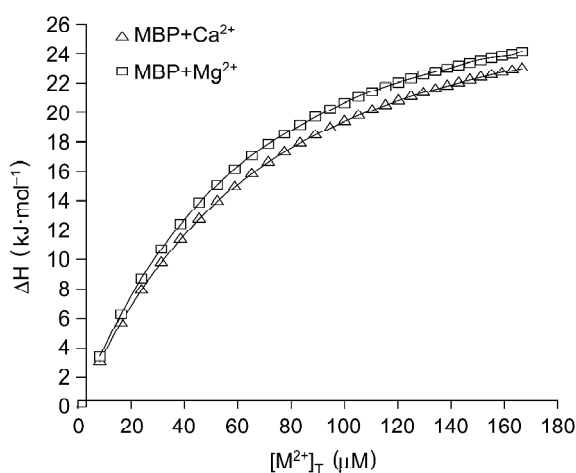


Fig. 1 Comparison between the experimental enthalpy values via **Equation 1** $[M^{2+}]_T$ is the total concentration of Ca^{2+} or Mg^{2+} ions in μM .

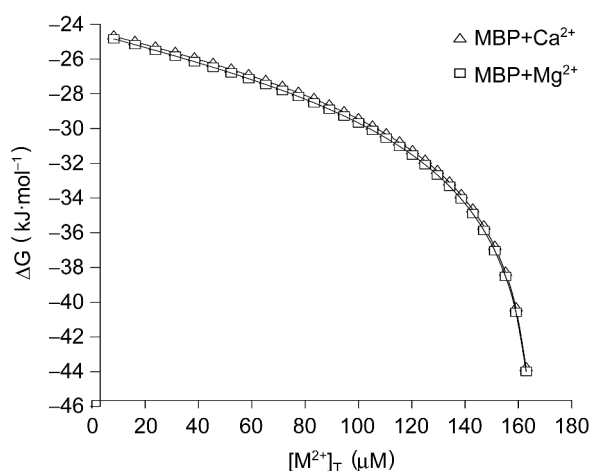


Fig. 2 Comparison between the experimental Gibbs free energies via **Equation 1** $[M^{2+}]_T$ is the total concentration of Ca^{2+} or Mg^{2+} ions in μM .

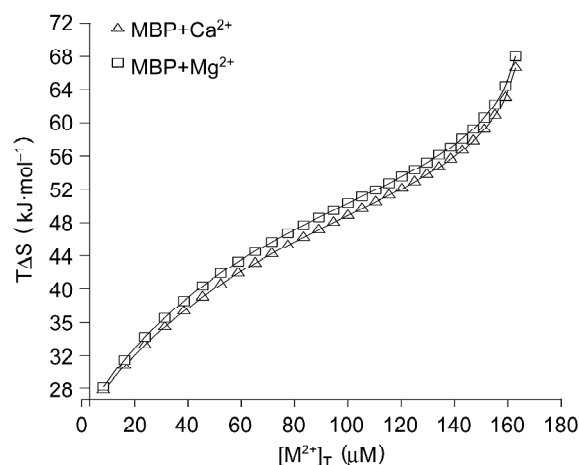


Fig. 3 Comparison between the experimental $T\Delta S$ values via **Equation 1** $[M^{2+}]_T$ is the total concentration of Ca^{2+} or Mg^{2+} ions in μM .

plexation with M^{2+} ions which can be expressed as follows:

$$\Phi = \frac{\Delta H}{\Delta H_{\max}} \quad 5$$

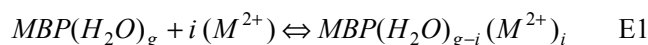
where ΔH_{dilut} represents the heat value upon saturation of all MBP. The appearance association equilibrium constant values, K_a , as a function of M^{2+} concentration can be calculated as follows:

$$K_a = \frac{\Phi}{(1 - \Phi)[M^{2+}]_F} \quad 6$$

$[M^{2+}]_F = (1 - x_B)[M^{2+}]_T$ is the unbounded or the free M^{2+} ion concentrations. The variable Φ represents the fraction of binding sites that are occupied on the peptide molecule. Therefore, $1 - \Phi$ represents the fraction of binding sites that are not occupied. The appearance association equilibrium constants, K_a , for successive replacement of water molecules by M^{2+} ions are as follow:

$$K_a = x_A^g - \sum_{i=1}^g K_i \frac{x_B^i}{x_A^{i-g}} \quad 7$$

where the K_i is the macroscopic association equilibrium constants which are the equilibrium constants for every successive replacement of water molecules by M^{2+} ions in the equilibria:



K_a values obtained from **Equation 6**, have been fitted to **Equation 7** using a computer program for nonlinear least-

Table 3 Binding parameters for MBP+Ca²⁺ and MBP+Mg²⁺ interactions calculated by Equations 1 and 7

	Ca ²⁺ +MBP	Mg ²⁺ +MBP
δ_A^o	0.053±0.016	0.968±0.100
δ_B^o	-12.322±0.046	-11.395±0.060
$K_1/\mu\text{M}^{-1}$	1.664±0.076	1.504±0.112
$K_2/\mu\text{M}^{-1}$	1.500±0.027	1.617±0.035
$\Delta H_{\text{max}}/\text{kJ}\cdot\text{mol}^{-1}$	23.000±0.007	24.152±0.038
g	2	2
p	1.040	1.040

$p=1$ indicates that the Ca²⁺ and Mg²⁺ ions occupy the identical binding sites non-cooperatively.

square fitting. Therefore, we can approach the “ g ” value and the macroscopic association equilibrium constant in the first, K_1 , and the second, K_2 , binding site (Table 3), which correspond to $\nu = 1$, $\nu = 2$ and $\nu = 3$ respectively. values can be calculated at any concentration of M²⁺ via Equation 2. The binding parameters obtained from this method are listed in Table 3. The Gibbs energies as a function of M²⁺ concentrations can be obtained as follow:

$$\Delta G = -RT \ln K_a \quad 8$$

Gibbs energies, ΔG , calculated from Equation 8 are shown graphically in Fig. 2. T ΔS values were calculated using ΔG values and have shown in Fig. 3. The less negative Gibbs free energies in the low M²⁺ concentrations (Fig. 2) indicate the lower affinity in this region.

A nonpolar residue dissolved in water induces a solvation shell in which water molecules are highly ordered. When two nonpolar groups come together on the folding of a polypeptide chain, the surface area exposed to the solvent is reduced, and part of the highly ordered water in the solvation shell is released to the bulk solvent, which results in an increase in the entropy. It is possible to introduce a correlation between change in δ_A^o and increase in the stability of proteins. The δ_A^o value reflects the hydrophobic property of MBP, leading to the enhancement of water structure. The greater the extent of this enhancement, the greater the stabilization of the MBP structure and the greater the value of δ_A^o . δ_A^o value (Table 3) for MBP+M²⁺ interactions are small and positive (Table 3), indicating that in the low concentration of M²⁺ the MBP structure is stabilized, resulting in an increase in its biological activity. δ_B^o value in high M²⁺ concentration are big and negative, indicating that the MBP structure is destabilized by these cations in this region.

The p values are very close to one ($p=1.04$), indicating

that there are a set of two identical non-operative binding sites for MBP+Ca²⁺ and MBP+Mg²⁺. In practice it turns out that even for good data it is usually very difficult to distinguish between heterogeneity and cooperativity, if more than two sites are present. Furthermore, it is almost impossible to derive accurate binding parameters from real data, if binding is cooperative and the binding sites are not equivalent.

A value of $p = 1$ would mean that $K_1 = K_2$, indicates that the binding of the second site occurs non-cooperatively as compared to binding of the first site, whereas a value of $p > 1$ would indicate that binding of the second site is facilitated and a value of $p < 1$ indicates that binding of the second site is inhibited (anti-cooperativity or negative cooperativity).

The same conclusion is reached via Equation 7 because the macroscopic association equilibrium constants recovered from this equation are roughly the same ($K_1 \approx K_2$ in Table 3).

Using fluorescence studies, Barylko *et al* have represented that there are two binding sites on MBP for Ca²⁺-calmodulin, but they could not determine the exact values of dissociation constants of every site [17]. Whereas, we can calculate all thermodynamic functions, cooperativity parameters, equilibrium constants and stability prediction as a result of ligand interaction with a biopolymer, just using Equations 1 and 7, and it is the most useful method in the ligand+macromolecule interactions.

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